



The Secret Life of Biofilms: How Microorganisms Build Their Own Cities

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ABSTRACT

Biofilms are highly organized, surface-attached microbial communities embedded within an extracellular polymeric matrix that enhance microbial survival, adaptability, and ecological success. Originating over 3.4 billion years ago, biofilms represent the dominant mode of microbial life and provide cells with protection from environmental stress, antimicrobial agents, and host immune responses. Biofilm development proceeds through sequential stages, initial attachment, irreversible adhesion, microcolony formation, maturation, and dispersion. Each regulated by complex molecular networks involving quorum sensing, second messengers such as c-di-GMP, and stress-response pathways. The biofilm mode of growth supports metabolic cooperation, efficient nutrient retention, and elevated horizontal gene transfer. Biofilms play critical roles across natural ecosystems, industrial systems, and clinical environments, contributing to both beneficial outcomes (e.g., wastewater treatment, bioremediation, plant growth promotion) and detrimental effects such as biofouling, corrosion, and persistent infections. Understanding the structural, molecular, and functional aspects of biofilms is essential for developing effective strategies for their control and harnessing their potential in environmental and biotechnological applications.

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INTRODUCTION

biofilm can be defined as a microbial community characterized by adhesion to a solid surface and by the production of an extracellular polymeric matrix in which the associated microorganism embedded (Alvarez-Ordonez et al., 2019). The **EPS** matrix typically consists polysaccharides, extracellular DNA (eDNA), lipids, proteins, and humic-like substances. This matrix forms a hydrated, gel-like scaffold that confers mechanical stability, facilitates nutrient retention, and protects embedded cells from external stresses.

Biofilm formation is considered an **adaptive developmental process**, involving regulated gene expression and coordinated microbial behaviour rather than simple aggregation.

1. HISTORY:

Biofilms originated around 3.4 billion years ago, when early microbes formed multispecies surface-attached communities to survive harsh environmental conditions. The first recorded observation of a biofilm was made by van Leeuwenhoek (1684),who described abundant microorganisms in dental plaque. Later, Heukelekian and Heller demonstrated the "bottle effect," showing that bacterial growth increases when surfaces are available. ZoBell confirmed that surface-attached cells occur in higher numbers than those in surrounding fluids. Advances in microscopy by Jones revealed the structural organization of biofilms, while Characklis showed that industrial biofilms are highly resistant to disinfectants such as chlorine. In 1978, Costerton and colleagues established the modern biofilm theory, recognizing biofilms as a predominant microbial lifestyle.

2. Stages of Biofilm Development

Biofilm formation generally progresses through five interrelated stages, although variations occur depending on species and environmental conditions

2.1 Initial Reversible Attachment

Planktonic cells encounter a surface through Brownian motion, hydrodynamic flow, or chemotactic movement. Weak physicochemical forces van Waals der interactions, hydrophobic contacts, electrostatic interactions allow temporary adhesion. Cell-surface appendages (flagella, pili, fimbriae) and adhesins enhance this interaction.

2.2 Irreversible Attachment

Adhering cells begin synthesizing EPS components, which anchor them firmly to the substrate. Changes in gene regulation often accompany this stage; for example, suppression of flagellar genes and induction of adhesin-encoding genes in bacteria such as *Pseudomonas aeruginosa*.

2.3 Microcolony Formation and EPS Accumulation

After stable attachment, the cells proliferate and recruit other microorganisms, forming microcolonies. EPS production increases significantly, creating a three-dimensional matrix. During this stage, the developing biofilm exhibits spatial heterogeneity, with nutrient and oxygen gradients forming across its depth (Giaouris et al., 2020).

2.4 Biofilm Maturation

Mature biofilms display **complex architecture**, often with mushroom-shaped structures, ridges, and interconnected water channels. These channels function as internal circulatory systems, distributing nutrients and removing metabolic waste. Microbial interactions, mutualism, competition, syntropy

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become pronounced, and the population may include multiple species.

2.5 Dispersion

In response to nutrient limitation, environmental signals, or quorum-sensing cues, subsets of cells detach from the biofilm. These dispersed cells regain motility and revert to the planktonic state, enabling colonization of new surfaces and continuation of the biofilm life cycle.

3. Molecular Regulation of Biofilm Formation

Biofilm development is tightly controlled by intricate regulatory networks:

3.1 Quorum Sensing

Quorum sensing (QS) involves production and detection of chemical signals (autoinducers). When cell density reaches a threshold, QS controls expression of genes linked to EPS synthesis, virulence, motility, and dispersion. Examples include:

- AHL (acyl-homoserine lactone) systems in Gram-negative bacteria
- Autoinducing peptides (AIPs) in Grampositive bacteria
- Furanosyl borate diester (AI-2) as a universal signal among diverse microbes

3.2 Second Messengers (e.g., c-di-GMP)

The cyclic di-GMP signalling pathway is central to microbial transition from motile to sessile lifestyles. High intracellular c-di-GMP levels promote EPS production, reduce motility, and stabilize biofilm structure.

3.3 Stress Response Systems: Environmental cues such as nutrient depletion, oxidative stress, or osmotic stress induce pathways—RpoS, two-component systems, and stringent

response factors that favour biofilm survival and resistance.

4. Benefits of Biofilm Mode of Growth

Microorganisms adopt the biofilm lifestyle for multiple advantages:

4.1 Increased Stress Tolerance

Biofilms offer exceptional protection against:

- antimicrobial agents
- desiccation
- heavy metals
- host immune responses

EPS layers slow diffusion of toxic compounds, while physiological heterogeneity creates subpopulations inherently more tolerant to stress.

4.2 Enhanced Genetic Exchange

Biofilms encourage horizontal gene transfer (HGT), including plasmid transfer and natural transformation, partly due to the presence of eDNA in the matrix.

4.3 Metabolic Cooperation

Microorganisms in biofilms engage in syntrophic relationships. For example, one species may produce metabolic by-products that serve as nutrients for another, enabling survival in nutrient-poor settings.

4.4 Resource Retention

EPS acts like a sponge, concentrating nutrients that would diffuse away in planktonic conditions.

- 5. Ecological and Applied Significance of Biofilms
- **5.1 Environmental Ecosystems:** Biofilms drive biogeochemical cycles, contribute to soil

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stability, and form primary colonizers in aquatic systems. They are crucial in nitrogen fixation, mineral weathering, and organic matter degradation.

- **5.2 Industrial Systems:** Biofilm formation can cause:
- biofouling in pipelines
- reduced efficiency in heat exchangers
- corrosion of metal surfaces
- contamination of food-processing equipment. These impacts carry significant economic costs.
- **5.3 Clinical Settings:** Biofilm-associated infections are particularly challenging due to enhanced antibiotic resistance. Examples include:
- · dental plaque
- chronic wound infections
- catheter-associated infections
- biofilms on implants and prosthetic devices

Cells in biofilms can survive antibiotic doses far higher than those effective against planktonic cells.

5.4 Beneficial Uses

Not all biofilms are harmful. They are essential in:

- wastewater treatment (activated sludge, trickling filters)
- bioremediation of pollutants
- plant-growth promotion through rootassociated biofilms
- 6. Approaches to Biofilm Control and Management: Control strategies aim to

prevent attachment, disrupt EPS, or interfere with regulatory systems:

- Anti-quorum sensing molecules to block communication
- **EPS-degrading enzymes** (DNases, glycosidases, proteases)
- Surface modifications to inhibit microbial adhesion
- Bacteriophages that target biofilm bacteria
- Nanoparticle-based antimicrobials
- **Physical approaches** such as shear stress, ultrasound, or surface abrasion

Emerging technologies seek to combine biochemical, physical, and genetic approaches for synergistic disruption of biofilms.

CONCLUSION

Biofilm formation sophisticated, is a multicellular developmental process that enhances microbial survival, stress resistance, and ecological fitness. Understanding biofilm architecture, molecular regulation, ecological roles remains a major area of microbiological research. Insights into biofilm biology not only help address biofilmassociated problems in medicine and industry but also enable the beneficial exploitation of microbial communities in environmental and agricultural systems.

REFERENCES

Alvarez-Ordóñez, A., Coughlan, L. M., Briandet, R., & Cotter, P. D. (2019). Biofilms in food processing environments: challenges and opportunities. *Annual Review of Food Science and Technology*, 10(1), 173-195.

Giaouris, E., Simões, M., & Dubois-Brissonnet, F. (2020). The role of biofilms in the development and dissemination of microbial resistance within the food industry. *Foods*, 9(6), 816.

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