

Lernaea – The Hidden Threat to Ornamental and Farmed Fish

Dhinesh P.*, Navin Kumar D., Porselvan S., Santhiya V., Kaviarasu D. and Saravanan S.

Dept. of Aquatic Animal Health Management, Tamil Nadu Dr. J. Jayalalithaa Fisheries University, Dr. M.G.R. Fisheries College and Research Institute, Ponneri, Tamil Nadu (601 204), India

Corresponding Author

Dhinesh P

Email: dhinesh1542@gmail.com



Lernaea, anchor worm, ornamental fish, aquaculture

How to cite this article:

Dhinesh, P., Kumar, N. D., Porselvan, S., Santhiya, V., Kaviarasu, D. and Saravanan, S. 2025. Lernaea – The Hidden Threat to Ornamental and Farmed Fish. *Vigyan Varta* 6 (11): 34-36.

ABSTRACT

Lernaea which is known as anchor worm exists as an ectoparasite that infects both ornamental and farmed freshwater fish species. The females attach themselves to skin and fins and gills to suck blood and tissue which leads to skin discomfort open wounds, stress, diminished growth and secondary infections. The organism produces free-swimming larvae which find hosts while its stationary females generate eggs during the entire time which results in fast population growth. The infestations appear as thread-like formations which makes them easy to identify. Management practices need to combine chemical applications with physical removal methods and water quality enhancement and fish welfare improvement.

INTRODUCTION

quaculture and the ornamental fish trade are rapidly expanding globally but parasite infestation is one of the limiting factors for sustainable production. Of these, *Lernaea*, (commonly referred to as the anchor worm), is a major external parasite of ornamental and farmed fish. This is a copepod

ectoparasite causing lernaeosis It belongs to the genus *Lernaea* of the Lernaeidae family (Hua *et al.*, 2019). The parasite anchors its head securely to all exterior parts of the fish, including some internal parts such as the mouth, gills, gill filaments, or even the eyes (Woo & Shariff, 1990). Upon attachment, it

November 2025 34 | P a g e



takes a blood meal and siphons host fish blood and body fluids resulting in irritation, lesions, and stress to the host fish. Lernaea infestation is unsightly not only in ornamental fishes, but also in aquaculture. The parasite debilitates fish, reduces growth and marketability and creates open sores that could be entry points for secondary bacterial and fungal infections. Heavy infection of lernaea may leads to high mortality in the case of young ones. The risks intensify because of environmental elements which include poor water quality together with large population densities and stressful conditions. The parasite Lernaea presents a hidden threat to fish health because it infects multiple hosts while spreading quickly and surviving well in water environments. Knowledge of its biology, significance, and control is important in the support of healthy stock in both the ornamental and commercial aquaculture industries.

Life Cycle of *Lernaea* – How It Grows and Spreads

The Lernaea (anchor worm) life cycle is both fascinating and detrimental to fish. The larvae start their development by passing through several stages which scientists call nauplius and copepodid until they find a fish host. The female worm transforms into a parasite after attachment where it extracts blood and tissue from the fish by inserting its head deep into skin or gills of fish. The females who remain anchored continue to produce eggs but the males remain unanchored and die soon after they reproduce. The cycle begins again when the eggs produce new larvae. The Lernaea parasite spreads quickly through aquaculture systems because it produces offspring continuously which makes control efforts extremely difficult.

Morphology of Lernaea

Lernaea shows a body structure that stands out from other species. The female stage of

fish exists as the most common stage which people commonly observe. Their body takes on a lengthy thread structure which usually spans between 5 and 12 millimetres. The front portion of the body contains a specialized anchor-shaped head structure cephalothorax which inserts deeply into the fish's body tissues. The parasite stays attached through this anchoring system while it feeds. The fish displays a clear trunk which extends from its head area. The parasite develops two lengthy egg sacs at its back end which form a "Y" shape. Males, in contrast, are much smaller, free-living, and rarely seen on fish.



Image Credit: the U.S. Geological Survey

Signs And Effects on Fish Health

Lernaea are visible with the naked eye; lernaea infestations are frequently simple to identify. The parasites show up as thin greenish-white threads which extend from the gills and fins and skin of the fish. The wound attachment sites develop inflammatory wounds and ulcers which produce red sores. Fish that infested frequently show signs restlessness while they use objects to relieve their discomfort. The parasite feeds on blood and bodily fluids which causes the host to lose strength while showing symptoms of reduced eating and stunted growth and body mass decrease. The condition will become worse and this will lead to bacterial and fungal infections in the affected area. Lernaea infestations threaten fish health and

November 2025 35 | P a g e



aquaculture production as they lead to anaemia and stress and sometimes result in fish death.

Host and Distribution

Lernaea is a common parasite which infects numerous freshwater fish species throughout the entire world. The parasite frequently in carp and goldfish and guppies and gouramis and catfish along with several other cultured and ornamental fish species. The parasite spreads easily through ponds and lakes and aquariums because it can live on various hosts. Warm climate conditions create an environment where infestations become common because these conditions enable pests to reproduce at a fast rate. The combination of high fish density in aquaculture ponds enables disease outbreaks to spread effectively. It has the ability to attack very broad range of hosts; this creates a severe problem for the both ornamental and farmed fishes.

Treatment and Control

Treatment and control of managing Lernaea requires both direct treatment of the parasite and improvement of the culture environment. The only way to get rid of individual parasites from aquarium fish requires manual extraction with tweezers. The common chemical treatments for ponds include potassium permanganate and formalin and salt baths. The medication organophosphates including trichlorfon undergo usage in regulated environments. (Noga, 2010). The treatment process requires precise execution because it must protect both fish health and water environmental cleanliness. The female Lernaea produces hundreds of eggs which makes it necessary to perform multiple treatment sessions. The control program

requires two essential components which include medication use and water quality improvement and fish stress reduction and proper hygiene maintenance.

CONCLUSION

Lernaea infestation creates a major problem for commercial and ornamental aquaculture because it damages fish health while slowing their growth and making them more vulnerable secondary opportunistic pathogens. Controlling oflernaea freshwater in environment is challenging due to its ability to infect multiple hosts and a quick offspring production rate within intensive fish culture. The management process requires early detection along with proper treatment, stress reduction and water quality improvement to succeed.

REFERENCE

Hua, C. J., Zhang, D., Zou, H., Li, M., Jakovlić, I., Wu, S. G., ... & Li, W. X. (2019). Morphology is not a reliable taxonomic tool for the genus *Lernaea*: molecular data and experimental infection reveal that *L. cyprinacea* and *L. cruciata* are conspecific. *Parasites* & vectors, 12(1), 579.

Noga, E. J. (2010). Fish disease: diagnosis and treatment. John Wiley & Sons.

Woo, P. T. K., & Shariff, M. (1990). Lernea cyprinacea L. (Copepoda: Caligidea) in Helostoma temmincki Cuvier & Valenciennes: the dynamics of resistance in recovered and naive fish. Journal of Fish Diseases, 13(6), 485-493.

November 2025 36 | Page